

Supplementary Information

A novel *Plasmodium falciparum* rhoptry associated adhesin mediates erythrocyte invasion through the sialic-acid dependent pathway

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Supplementary Methods

Mass Spectrometric analysis of purified recombinant protein

In-gel digestion of the purified recombinant protein band was done to ascertain its identity. The corresponding protein bands from the coomassie stained SDS-PAGE gel were excised, reduced, and alkylated. Proteins were digested overnight with trypsin (Promega) and the corresponding peptides were eluted from the gel using trifluoroacetic acid. Mass spectrometry analysis and protein identification was done as described previously¹

ELISA

Antibody responses in mice and rabbit were quantified by ELISA. Briefly, 96-well plates (Costar) were coated overnight with 0.2 µg per well of the recombinant protein in 0.06 M carbonate-bicarbonate buffer, pH 9.6 (Sigma). The plates were washed thrice with 0.05% Tween in PBS and further blocked with 2% skimmed milk in phosphate buffered saline (PBS) at 37°C for 2 h. Washing was sequentially repeated and serial dilutions of the primary sera (1:1,000-fold onwards) were prepared and incubated in the respective wells for 1 hour. The ELISA plate was subjected to stringent washing with PBS containing 0.05% Tween-20 and finally with PBS alone. Thereafter, a 1:10,000 dilution of the horseradish peroxidase conjugated secondary antibody (Sigma, St. Louis, MO) was added to each well and incubated for 60 minutes at 37°C. The enzymatic reaction was developed by the addition of *o*-phenylenediamine dihydrochloride (OPD) and hydrogen peroxide for 25 minutes at 37°C. The reaction was terminated by the addition of 2M sulphuric acid, and OD₄₉₂ was recorded by using an ELISA microplate reader (Molecular Devices). Pre-immune (pre-bleed; PI) sera were used at similar dilutions as a control.

FACS-based erythrocyte binding assay (EBA)

Erythrocytes were incubated with 0.5 ml of culture supernatant at 37°C for 3 hours. After incubation, the erythrocytes were washed with PBS and bound PfRA on surface of erythrocytes was detected using anti-PfRA rabbit sera and Alexa 488 conjugated goat anti-rabbit IgG antibodies and were read on a FACS Caliber flow cytometer. The resulting flow cytometry data were analyzed using Cell Quest software.

Co-Immunoprecipitation and Mass Spectrometric Analysis

Immunoprecipitation experiments using lysate of the schizont stage parasites were performed as prescribed (Thermo scientific). The trypsin digested samples were analyzed on a nano-LC equipped Orbitrap VELOS PRO (Thermo Fisher Scientific) mass spectrometer as described previously¹. The proteins were identified by blasting the peptides over a *Plasmodium falciparum* database (Uniprot), using proteome discoverer (Thermofisher) using standardized procedure as described earlier¹.

Invasion assays

Invasion assays were done as described previously². Briefly, the parasites were first synchronized by the purification of schizont-stage (40-42 h) parasites on a percoll gradient. 0.4 million schizont stage percoll gradient purified parasites were incubated with 20 million uninfected normal as well as enzymatically treated erythrocytes at 2% haematocrit. After about 40 h post invasion, the parasite infected erythrocytes were stained with ethidium bromide dye and measured by a fluorescence activated cell sorter (FACS)-based assay. Relative invasion in enzyme treated erythrocytes was calculated with respect to the invasion observed in untreated erythrocytes. Two different experiments were performed in triplicate. For each assay Dd2 was

used as control for neuraminidase treated erythrocytes². The error bars represent the standard error of the mean.

Sequence polymorphism analysis

To check sequence polymorphisms in the PfRA gene, available nucleotide sequences for the *P. falciparum* laboratory clones and field isolates was retrieved from the Plasmodb database, translated using ExPASy translate tool and aligned using multiple sequence alignment program Clustal Omega.

References

1. Reddy, K. S. *et al.* Multiprotein complex between the GPIanchored CyRPA with PfRH5 and PfRipr is crucial for *Plasmodium falciparum* erythrocyte invasion. *Proc. Natl. Acad. Sci. U.S.A.* **112**, 1179–1184 (2015).
2. Gaur, D., Storry, J.R., Reid, M.E., Barnwell, J.W., Miller, L.H. *Plasmodium falciparum* Is Able To Invade Erythrocytes through a Trypsin-Resistant Pathway Independent of Glycophorin B. *Infect Immun.* **71**, 6742-6746 (2003).

Supplementary Figure S1: Alignment of PfRA protein sequences from 9 *P. falciparum* laboratory strains (Clustal omega).

3D7	MKRFVLFVIFLVHIWSENVDTFKCNYSKKKNGHHIKRHTNDEEKKEYSFLMLGKEN	60
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Dd2-2	MKRFVLFVIFLVHIWSENVDTFKCNYSKKKNGHHIKRHTNDEEKKEYSFLMLGKEN	
7G8	MKRFVLFVIFLVHIWSENVDTFKCNYSKKKNGHHIKRHTNDEEKKEYSFLMLGKEN	
IT	MKRFVLFVIFLVHIWSENVDTFKCNYSKKKNGHHIKRHTNDEEKKEYSFLMLGKEN	
707A	MKRFVLFVIFLVHIWSENVDTFKCNYSKKKNGHHIKRHTNDEEKKEYSFLMLGKEN	
HB3	MKRFVLFVIFLVHIWSENVDTFKCNYSKKKNGHHIKRHTNDEEKKEYSFLMLGKEN	
CS2	MKRFVLFVIFLVHIWSENVDTFKCNYSKKKNGHHIKRHTNDEEKKEYSFLMLGKEN	

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Dd2-2	EENKENKENQNVNPKDNNDNNNEKKNEEQHNNEKKQEEVINNNNNNVENKKEEENHN	
7G8	EENKENKENQNVNPKDNNDNNNEKKNEEQHNNEKKQEEVINNNNNNVENKKEEENHN	
IT	EENKENKENQNVNPKDNNDNNNEKKNEEQHNNEKKQEEVINNNNNNVENKKEEENHN	
707A	EENKENKENQNVNPKDNNDNNNEKKNEEQHNNEKKQEEVINNNNNNVENKKEEENHN	
HB3	EENKENKENQNVNPKDNNDNNNEKKNEEQHNNEKKQEEVINNNNNNVENKKEEENHN	
CS2	EENKENKENQNVNPKDNNDNNNEKKNEEQHNNEKKQEEVINNNNNNVENKKEEENHN	

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T9_94	NDKKTDEQNHVNEKKOEGENNKKDDTIPPEKKINKENLLEYGTHDKEGHFIPSYKTLT	
Dd2-2	NDKKTDEQNHVNEKKOEGENNKKDDTIPPEKKINKENLLEYGTHDKEGHFIPSYKTLT	
7G8	NDKKTDEQNHVNEKKOEGENNKKDDTIPPEKKINKENLLEYGTHDKEGHFIPSYKTLT	
IT	NDKKTDEQNHVNEKKOEGENNKKDDTIPPEKKINKENLLEYGTHDKEGHFIPSYKTLT	
707A	NDKKTDEQNHVNEKKOEGENNKKDDTIPPEKKINKENLLEYGTHDKEGHFIPSYKTLT	
HB3	NDKKTDEQNHVNEKKOEGENNKKDDTIPPEKKINKENLLEYGTHDKEGHFIPSYKTLT	
CS2	NDKKTDEQNHVNEKKOEGENNKKDDTIPPEKKINKENLLEYGTHDKEGHFIPSYKTLT	

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HB3	DEILSTNNNAMEKASSFLKIACSHVMKLIEFIPESKLSSQYIKVDNKNIYLKDIAVECQNI	
CS2	DEILSTNNNAMEKASSFLKIACSHVMKLIEFIPESKLSSQYIKVDNKNIYLKDIAVECQNI	

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HB3	YFNLEKFSMTLIVFNSKINKIFIYSQEK	
CS2	YFNLEKFSMTLIVFNSKINKIFIYSQEK	

Supplementary Figure S2: Alignment of PfRA protein sequences from 57 *P. falciparum* field isolates
(Clustal omega).

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309.1	MKRFFVLFVIFLVHIWSENVDTFKCNYSKKKNGHHIKRHITNDEEKKEYSFLMLGKEN	
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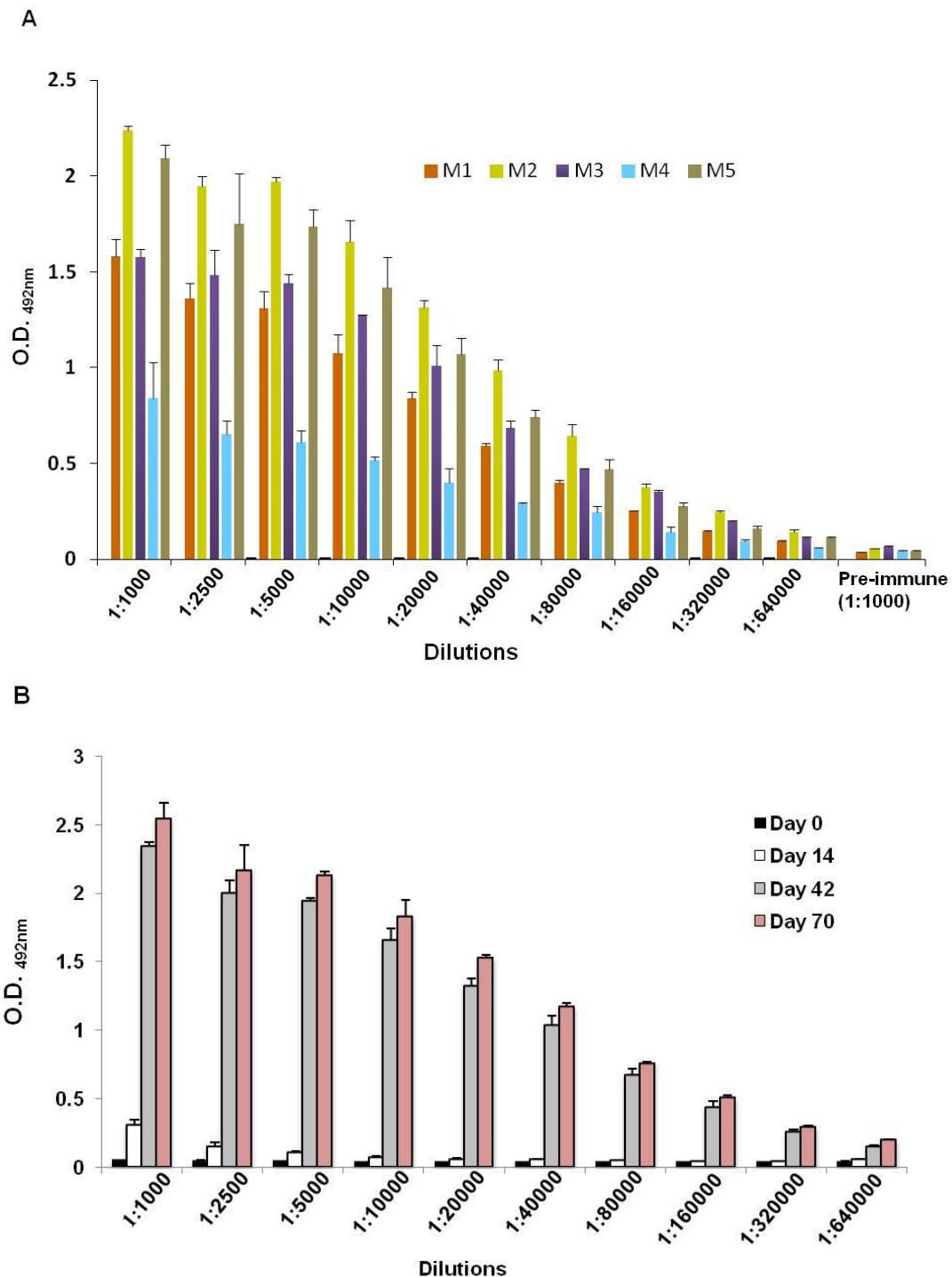
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SentT149_09	NDKKTDEQNHVNEKKOEGENNKKDDTIPPEKKINKENLLEYGTHDKEGHFIPSYKTLT	
SentT180_08	NDKKTDEQNHVNEKKOEGENNKKDDTIPPEKKINKENLLEYGTHDKEGHFIPSYKTLT	
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RV_3714	NDKKTDEQNHVNEKKOEGENNKKDDTIPPEKKINKENLLEYGTHDKEGHFIPSYKTLT	
RV_3717	NDKKTDEQNHVNEKKOEGENNKKDDTIPPEKKINKENLLEYGTHDKEGHFIPSYKTLT	
327_1	NDKKTDEQNHVNEKKOEGENNKKDDTIPPEKKINKENLLEYGTHDKEGHFIPSYKTLT	
RV_3655	NDKKTDEQNHVNEKKOEGENNKKDDTIPPEKKINKENLLEYGTHDKEGHFIPSYKTLT	
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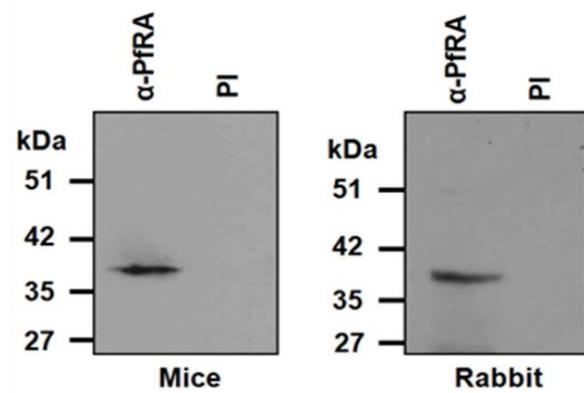
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RV_3637	DEILSTNNNAMEKASSFLKIA CSHVMKLIEFIPESKLSSQYIKVDNKNIYLKDIAVECQNI	
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RV_3606	DEILSTNNNAMEKASSFLKIA CSHVMKLIEFIPESKLSSQYIKVDNKNIYLKDIAVECQNI	
PS103	DEILSTNNNAMEKASSFLKIA CSHVMKLIEFIPESKLSSQYIKVDNKNIYLKDIAVECQNI	
RV_3741	DEILSTNNNAMEKASSFLKIA CSHVMKLIEFIPESKLSSQYIKVDNKNIYLKDIAVECQNI	
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N011-A	DEILSTNNNAMEKASSFLKIA CSHVMKLIEFIPESKLSSQYIKVDNKNIYLKDIAVECQNI	
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M113-A	DEILSTNNNAMEKASSFLKIA CSHVMKLIEFIPESKLSSQYIKVDNKNIYLKDIAVECQNI	
RV_3714	DEILSTNNNAMEKASSFLKIA CSHVMKLIEFIPESKLSSQYIKVDNKNIYLKDIAVECQNI	
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327.1	DEILSTNNNAMEKASSFLKIA CSHVMKLIEFIPESKLSSQYIKVDNKNIYLKDIAVECQNI	
RV_3655	DEILSTNNNAMEKASSFLKIA CSHVMKLIEFIPESKLSSQYIKVDNKNIYLKDIAVECQNI	
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TRIPS_480	YFNLEKFSMTLIVFNSKINKFIYSQEK	
RV_3703	YFNLEKFSMTLIVFNSKINKFIYSQEK	
RV_3637	YFNLEKFSMTLIVFNSKINKFIYSQEK	
RV_3729	YFNLEKFSMTLIVFNSKINKFIYSQEK	
RV_3708	YFNLEKFSMTLIVFNSKINKFIYSQEK	
UGK_408.2	YFNLEKFSMTLIVFNSKINKFIYSQEK	
309.1	YFNLEKFSMTLIVFNSKINKFIYSQEK	
UGK_443.2	YFNLEKFSMTLIVFNSKINKFIYSQEK	
RV_3736	YFNLEKFSMTLIVFNSKINKFIYSQEK	
PS122_G11	YFNLEKFSMTLIVFNSKINKFIYSQEK	
RV_3687	YFNLEKFSMTLIVFNSKINKFIYSQEK	
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PS103	YFNLEKFSMTLIVFNSKINKFIYSQEK	
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TRIPS_364	YFNLEKFSMTLIVFNSKINKFIYSQEK	
BM_0009	YFNLEKFSMTLIVFNSKINKFIYSQEK	
TRIPS_331	YFNLEKFSMTLIVFNSKINKFIYSQEK	
RV_3766	YFNLEKFSMTLIVFNSKINKFIYSQEK	
TRIPS_355	YFNLEKFSMTLIVFNSKINKFIYSQEK	
TRIPS_461	YFNLEKFSMTLIVFNSKINKFIYSQEK	
TRIPS_501	YFNLEKFSMTLIVFNSKINKFIYSQEK	
O079-B	YFNLEKFSMTLIVFNSKINKFIYSQEK	
H209	YFNLEKFSMTLIVFNSKINKFIYSQEK	
P196J3-C	YFNLEKFSMTLIVFNSKINKFIYSQEK	
SenT101.09	YFNLEKFSMTLIVFNSKINKFIYSQEK	
SenT149.09	YFNLEKFSMTLIVFNSKINKFIYSQEK	
SenT180.08	YFNLEKFSMTLIVFNSKINKFIYSQEK	
SenT142.09	YFNLEKFSMTLIVFNSKINKFIYSQEK	
N011-A	YFNLEKFSMTLIVFNSKINKFIYSQEK	
O306-A	YFNLEKFSMTLIVFNSKINKFIYSQEK	
P241-D	YFNLEKFSMTLIVFNSKINKFIYSQEK	
RV_3635	YFNLEKFSMTLIVFNSKINKFIYSQEK	
P164-C	YFNLEKFSMTLIVFNSKINKFIYSQEK	
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RV_3739	YFNLEKFSMTLIVFNSKINKFIYSQEK	
RV_3730	YFNLEKFSMTLIVFNSKINKFIYSQEK	
BM_0008	YFNLEKFSMTLIVFNSKINKFIYSQEK	
TRIPS_759	YFNLEKFSMTLIVFNSKINKFIYSQEK	
RV_3735	YFNLEKFSMTLIVFNSKINKFIYSQEK	
TRIPS_440	YFNLEKFSMTLIVFNSKINKFIYSQEK	
RV_3721	YFNLEKFSMTLIVFNSKINKFIYSQEK	
P237-C	YFNLEKFSMTLIVFNSKINKFIYSQEK	
TRIPS_487	YFNLEKFSMTLIVFNSKINKFIYSQEK	
M113-A	YFNLEKFSMTLIVFNSKINKFIYSQEK	
RV_3714	YFNLEKFSMTLIVFNSKINKFIYSQEK	
RV_3717	YFNLEKFSMTLIVFNSKINKFIYSQEK	
327.1	YFNLEKFSMTLIVFNSKINKFIYSQEK	
RV_3655	YFNLEKFSMTLIVFNSKINKFIYSQEK	

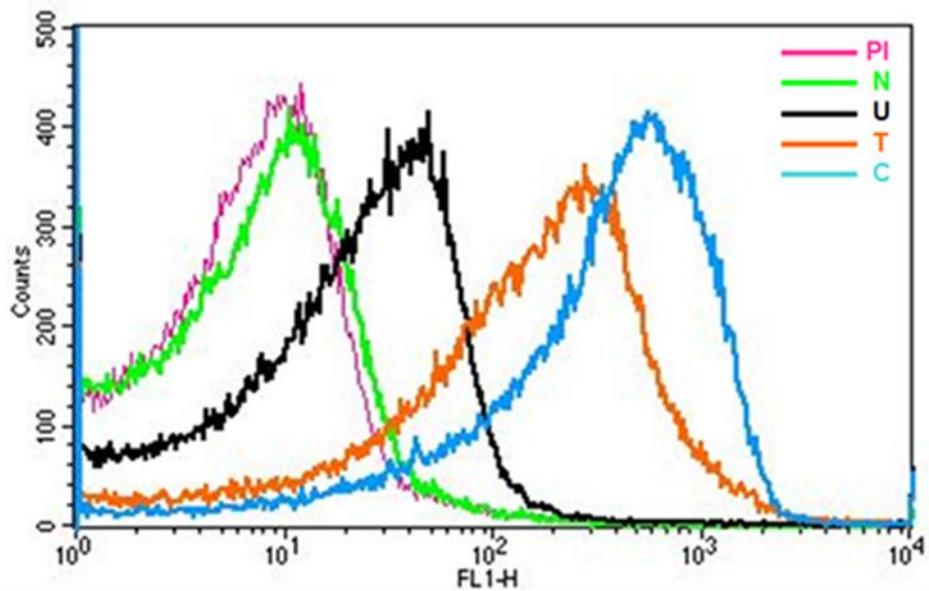
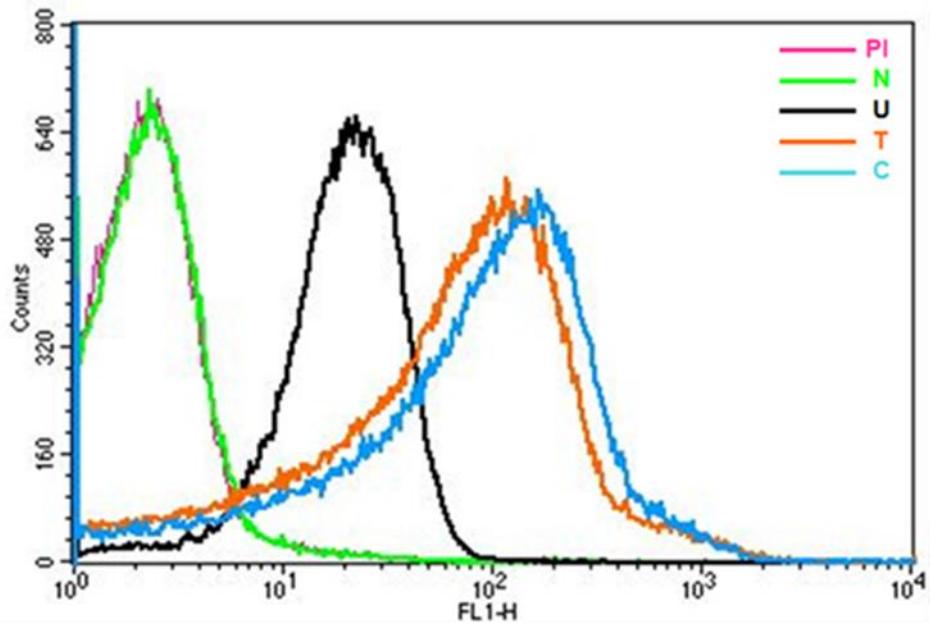
Supplementary Figure S3: Measurement of the antibody responses (end point titers) against rPfRA. Immunogenicity of rPfRA in (A) mice and (B) rabbit was analyzed by ELISA. Sera were serially diluted and assessed for end point titers. Pre-immune sera were taken as controls. High titer antibodies (end point observed at dilution of 1:320,000 in mice and 1:640,000 in rabbit) against the recombinant PfRA protein were detected. The error bars represent the standard error of the mean.



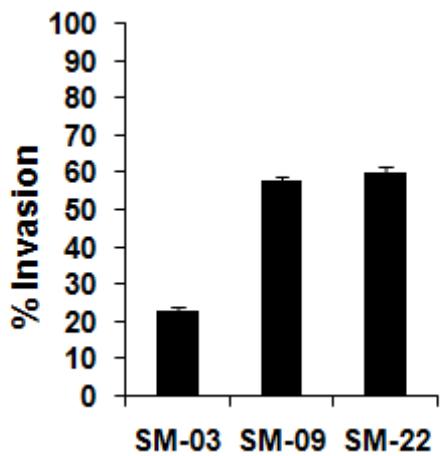
Supplementary Figure S4: Immunoblot analysis of rPfRA with immune sera raised in mice and rabbit. I: immune sera; PI: pre-immune sera.



Supplementary Figure S5: Binding of (A) native PfRA and (B) rPfRA to untreated (U), neuraminidase-treated (N), trypsin-treated (T) and chymotrypsin-treated (C) human erythrocytes was detected by flow cytometry using anti-PfRA rabbit sera. Pre-immune rabbit sera (PI) was used as a control.

A**B**

Supplementary Figure S6: Invasion phenotype of three freshly cultured *P. falciparum* field isolates from Manhica, Mozambique in neuraminidase-treated erythrocytes.



Supplementary Table S1: Geographical origin of *P. falciparum* laboratory clones (www.plasmodb.org).

No.	<i>P. falciparum</i> laboratory clone	Geographical location
1.	3D7	Africa
2.	Dd2-1	Laos, Indochina
3.	T9_94	Thailand
4.	Dd2-2	Laos, Indochina
5.	7G8	Brazil
6.	IT	Brazil
7.	707A	Unknown
8.	HB3	Honduras
9.	CS2	Brazil

Supplementary Table S2: Geographical origin of *P. falciparum* field isolates
(www.plasmodb.org).

No.	<i>P. falciparum</i> field isolate	Geographical location
1.	TRIPS_480	Gambia
2.	RV_3703	Gambia
3.	RV_3637	Gambia
4.	RV_3729	Gambia
5.	RV_3708	Gambia
6.	UGK_408.2	Uganda, Kampala
7.	309.1	Mali
8.	UGK_443.2	Uganda, Kampala
9.	RV_3736	Gambia
10.	PS122_G11	Mali, Bandiagara
11.	RV_3687	Gambia
12.	RV_3610	Gambia
13.	PS250	Mali, Bandiagara
14.	RV_3675	Gambia
15.	RV_3606	Gambia
16.	PS103	Mali, Bandiagara
17.	RV_3741	Gambia
18.	TRIPS_437	Gambia
19.	TRIPS_474	Gambia
20.	RV_3701	Gambia
21.	PS183	Mali, Bandiagara
22.	RV_3731	Gambia
23.	TRIPS_364	Gambia
24.	BM_0009	Gambia
25.	TRIPS_331	Gambia
26.	RV_3766	Gambia
27.	TRIPS_355	Gambia
28.	TRIPS_461	Gambia
29.	TRIPS_501	Gambia
30.	O079-B	French Guiana, Cayenne
31.	H209	French Guiana
32.	P196J3-C	Saint-Laurent du Maroni
33.	SenT101.09	Senegal,Thies
34.	SenT149.09	Senegal,Thies
35.	SenT180.08	Senegal,Thies
36.	SenT142.09	Senegal,Thies
37.	N011-A	French Guiana, Saul
38.	O306-A	French Guiana, Cayenne
40.	RV_3635	Gambia
41.	P164-C	French Guiana, Cayenne

No.	<i>P. falciparum</i> field isolate	Geographical location
42.	RV_3630	Gambia
43.	RV_3739	Gambia
44.	RV_3730	Gambia
45.	BM_0008	Gambia
46.	TRIPS_759	Gambia
47.	RV_3735	Gambia
49.	TRIPS_440	Gambia
50.	RV_3721	Gambia
51.	P237-C	French Guiana, Cayenne
52.	TRIPS_487	Gambia
53.	M113-A	Saint Georges de l'Oyapock
54.	RV_3714	Gambia
55.	RV_3717	Gambia
56.	327.1	Mali
57.	RV_3655	Gambia

Supplementary Table S3: List of Unique Peptides generated from the Mass Spectrometric analysis (LC-MS) of the trypsin digested recombinant PfRA protein.

Accession	Description	Score	Coverage
Q8IJS3	Conserved Plasmodium protein OS=Plasmodium falciparum (isolate 3D7) GN=PF10_0119 PE=4 SV=1 - [Q8IJS3_PLAF7]	1226.62	79.03
	A2	Sequence of Unique Peptides	# PSMs
	High	INKENLLEYGTHDKEGHFIPSYK	50
	High	ENEENKENKENNQNVNK	4
	High	ENLLEYGTHDKEGHFIPSYK	31
	High	KQEEVINNNNNNVENKK	5
	High	EYSFLMLGKENEEENKENK	8
	High	EYSFLmLGKENEEENKENK	16
	High	IAcSHVmKLIEFIPESK	2
	High	LIEFIPESKLSSQYIK	5
	High	KINKENLLEYGTHDK	11
	High	KQEEVINNNNNNVENK	6
	High	EYSFLMLGKENEEENK	3
	High	QEEVINNNNNNVENK	2
	High	KDDTIPPEKK	18
	High	INKENLLEYGTHDK	12
	High	DIAVEcQNIYFNLEK	26
	High	TLTDEILSTNNAMEK	25
	High	EYSFLmLGKENEEENK	11
	High	KNEEQHNNEK	18
	High	KTDEQNHVNEK	5
	High	TDEQNHVNEK	12
	High	TLTDEILSTNNAmEK	60
	High	VDNKNIYLK	3
	High	KKEYSFLmLGK	8
	High	ENLLEYGTHDK	16
	High	KTDEQNHVNEKK	1
	High	HITNDEEKKK	14

Supplementary Table S4: Mass spectrometric identification of native PfRA from immunoprecipitation elutes using specific antibodies.

Accession	Description	Score	Coverage	Unique Peptides
PF3D7_1012200	Organism = Plasmodium_falciparum_3D7 product = conserved Plasmodium protein, unknown function location = Pf3D7_10_v3:470979 - 471933(+) length=267 sequence_SO = chromosome SO=protein_coding	44.32	22.85	TLTDEILSTnNAMEK TLTDEILSTNNAMEK DIAVECQNIYFNLEK TLTDEILSTNNAmEK DIAVEcQNIYFNLEK FSMTLIVFNSK LIEFIPESK HKKDDTIPPEK FSMTLIVFnSK

PfRA protein sequence: Unique peptides detected by LC-MS highlighted in green

Signal Sequence (1-22) →
MKRFFVLVIFLVHIWSENVDTFKCNYSKKKKNGHHIKRHITNDEEKKEYSFLMLGKE
 NEEENKENKENQNQNVNKDNNDNNNNEKKNEEQNHNEKKQEEVINNNNNVENKKE
 EENHNNDKKDEQNHVNEKKQEGENNKHKKDDTIPPEK KINKENLLEYGTHDKEGHFIP
 SYK TLTDEILSTNNAMEK ASSFLKIACSHVMK LIEFIPESKLSSQYIKVDNKNIYLKDIAVE
 CQNIYFNLEKFSMTLIVFNSK INKFIYSQEK